A Focus towards p-vinyl benzaldehyde containing quinoxaline - Photoluminescence and Antibacterial activity

R. Padma, S. Guhanathan*
PG & Research Department of Chemistry,
Muthurangam Government Arts College (Autonomous), Vellore, India
Corresponding Author: R. Padma

ABSTRACT: Quinoxaline display a wide range of biological performance and electron transport properties. Poly(p-phenylene vinylene) is one of the most imperative part of conjugated polymers having an ample assortment of applications in light-emitting diodes. The existence of electron withdrawing quinoxaline ring has been used in π-conjugated structures to construct the OLED materials. In the present investigation, p-vinyl benzaldehyde substituted quinoxaline derivatives were synthesized using 3-methyl-quinoxalin-2-one with terephthal dicarboxaldehyde via Wittig reaction. The structures of synthesised compounds were confirmed by FT-IR, 1H, 13C, 31P-NMR, MASS spectral data. The result of Fluorescent investigational examination reveals that p-vinyl benzaldehyde containing quinoxaline derivatives exhibited photoluminescence nature with green emission maxima at shorter wavelengths of 470nm. The Phosphonium compound and p-vinyl benzaldehyde capped quinoxaline derivatives were subjected to four different bacteria viz., Staphylococcus aureus, Bacillus subtilis, Escherichia Coli, Pseudomonas auruginosa. A result of the antibacterial studies reveals that compounds acquire substantial activity in assessment with Ampicillin.

Keywords: p-phenylene vinylene, quinoxaline derivatives, photoluminescence, Wittig reaction, spectral studies.

I. INTRODUCTION

Quinoxaline derivatives are an important class of fused heterocycles that display a wide range of biological, pharmacological, and medicinal properties involving antiviral, antibacterial, anti-inflammatory, and anti-protozoal and as kinase inhibitors[1-5]. Many quinoxaline derivatives have a wide application as dyes, electroluminescent materials, organic semiconductors, cavitarinds, chemically controllable switches, and DNA cleaving agents [6-11]. The conjugated polymers have attracted significant attraction due to their vast applications in numerous fields of research viz., photodiodes[12], photovoltaic cells [13], nonlinear optics[14], and laser devices [15]. The emergence of conjugated polymers as a new class of electronic materials has attracted many researchers for their tremendous potential applications. Conjugated polymers are semiconductors viz., poly(phenylene vinylene)s (PPVs), polyfluorenes(PFs), polythiophene (PT) have been synthesized and utilized as emissive layers in light-emitting diodes (LED) [16], p-type (electron donor, hole transport) and n-type (electron acceptor, electron transport) based polymeric semiconductors are essentially needed for creating more efficient and high performance electronic and opto-electronic devices[17]. Among the types of polymeric semi-conductors, n-type semi-conductors are feasible with nitrogen heterocyclic containing polymers jointly considered as “heterocyclic polymers”.

In general, the conjugated polymers, the barrier of electron injection is much higher than that of hole injection. To improve the efficiency of these, it is necessary to balance the rate of injection of electrons and holes from opposite electrodes into the device. Hence, the high electron affinity substituents such as quinoxaline [18-20], oxadiazole[21], triazole[22], and quinoline[23] have been introduced into conjugated polymers. It has been well known that inclusion of heterocyclic compound in the back of conjugated polymers found to have good semi-conducting properties with outstanding thermal and oxidative stability, low moisture absorbing capabilities and excellent film forming capacities. Although, a variety of N-heterocyclic compounds containing PPV viz, oxadiazole, pyridine, quinoline, substituted quinoline, have been published[24-25].

To our knowledge, there is not much report apart from our group on substituted quinoxaline containing PPV. Further, it would be an interesting to investigate the effect of substitutent on the optical, electronic properties of conjugated polymer. In this paper, our interest towards the synthesis, photophysical and electrochemical aspects of a new conjugated vinyl benzaldehyde capped with quinoxaline derivative.
II. EXPERIMENTAL

2.1. Materials
All the chemicals were obtained from Avra chemicals, Hyderabad, India and were used as supplied. Solvents used were purified and dried according to the standard procedure.

2.2. Characterisation Methods
The UV-Visible spectra were recorded on an Alpha-Bruker UV spectrophotometer equipped in the range between 200-800nm. Room temperature FTIR spectra were recorded as KBr pellet with an Alpha-Bruker FTIR spectrophotometer in the range of 4000-400cm⁻¹. Nuclear magnetic resonance spectra with different core viz., ¹H NMR, ¹³C NMR and ³¹P NMR were recorded in either DMSO-d₆ or CDCl₃ on Bruker ADVANCE III 500mHz spectrometer. The fluorescence spectra of the synthesised compound in ethanol were recorded on fluorescence spectrophotometer, FP-8500, JASCO. Mass spectroscopy was recorded on ES-FIGIEAN ionization mass spectrometer.

Fig. 3419.79(N chromatography using silica gel. Hexane

2.2.1. Materials
Solvents used were purified and dried according to the standard procedure.

2.2.2. Characterisation Methods
The UV-Visible spectra were recorded on an Alpha-Bruker UV spectrophotometer equipped in the range between 200-800nm. Room temperature FTIR spectra were recorded as KBr pellet with an Alpha-Bruker FTIR spectrophotometer in the range of 4000-400cm⁻¹. Nuclear magnetic resonance spectra with different core viz., ¹H NMR, ¹³C NMR and ³¹P NMR were recorded in either DMSO-d₆ or CDCl₃ on Bruker ADVANCE III 500mHz spectrometer. The fluorescence spectra of the synthesised compound in ethanol were recorded on fluorescence spectrophotometer, FP-8500, JASCO. Mass spectroscopy was recorded on ES-FIGIEAN ionization mass spectrometer.

Fig. 3419.79(N chromatography using silica gel. Hexane

2.2.1. Materials
Solvents used were purified and dried according to the standard procedure.

2.2.2. Characterisation Methods
The UV-Visible spectra were recorded on an Alpha-Bruker UV spectrophotometer equipped in the range between 200-800nm. Room temperature FTIR spectra were recorded as KBr pellet with an Alpha-Bruker FTIR spectrophotometer in the range of 4000-400cm⁻¹. Nuclear magnetic resonance spectra with different core viz., ¹H NMR, ¹³C NMR and ³¹P NMR were recorded in either DMSO-d₆ or CDCl₃ on Bruker ADVANCE III 500mHz spectrometer. The fluorescence spectra of the synthesised compound in ethanol were recorded on fluorescence spectrophotometer, FP-8500, JASCO. Mass spectroscopy was recorded on ES-FIGIEAN ionization mass spectrometer.

Fig. 3419.79(N chromatography using silica gel. Hexane

2.2.1. Materials
Solvents used were purified and dried according to the standard procedure.

2.2.2. Characterisation Methods
The UV-Visible spectra were recorded on an Alpha-Bruker UV spectrophotometer equipped in the range between 200-800nm. Room temperature FTIR spectra were recorded as KBr pellet with an Alpha-Bruker FTIR spectrophotometer in the range of 4000-400cm⁻¹. Nuclear magnetic resonance spectra with different core viz., ¹H NMR, ¹³C NMR and ³¹P NMR were recorded in either DMSO-d₆ or CDCl₃ on Bruker ADVANCE III 500mHz spectrometer. The fluorescence spectra of the synthesised compound in ethanol were recorded on fluorescence spectrophotometer, FP-8500, JASCO. Mass spectroscopy was recorded on ES-FIGIEAN ionization mass spectrometer.

Fig. 3419.79(N chromatography using silica gel. Hexane

2.2.1. Materials
Solvents used were purified and dried according to the standard procedure.

2.2.2. Characterisation Methods
The UV-Visible spectra were recorded on an Alpha-Bruker UV spectrophotometer equipped in the range between 200-800nm. Room temperature FTIR spectra were recorded as KBr pellet with an Alpha-Bruker FTIR spectrophotometer in the range of 4000-400cm⁻¹. Nuclear magnetic resonance spectra with different core viz., ¹H NMR, ¹³C NMR and ³¹P NMR were recorded in either DMSO-d₆ or CDCl₃ on Bruker ADVANCE III 500mHz spectrometer. The fluorescence spectra of the synthesised compound in ethanol were recorded on fluorescence spectrophotometer, FP-8500, JASCO. Mass spectroscopy was recorded on ES-FIGIEAN ionization mass spectrometer.

Fig. 3419.79(N chromatography using silica gel. Hexane

2.2.1. Materials
Solvents used were purified and dried according to the standard procedure.

2.2.2. Characterisation Methods
The UV-Visible spectra were recorded on an Alpha-Bruker UV spectrophotometer equipped in the range between 200-800nm. Room temperature FTIR spectra were recorded as KBr pellet with an Alpha-Bruker FTIR spectrophotometer in the range of 4000-400cm⁻¹. Nuclear magnetic resonance spectra with different core viz., ¹H NMR, ¹³C NMR and ³¹P NMR were recorded in either DMSO-d₆ or CDCl₃ on Bruker ADVANCE III 500mHz spectrometer. The fluorescence spectra of the synthesised compound in ethanol were recorded on fluorescence spectrophotometer, FP-8500, JASCO. Mass spectroscopy was recorded on ES-FIGIEAN ionization mass spectrometer.

Fig. 3419.79(N chromatography using silica gel. Hexane

2.2.1. Materials
Solvents used were purified and dried according to the standard procedure.

2.2.2. Characterisation Methods
The UV-Visible spectra were recorded on an Alpha-Bruker UV spectrophotometer equipped in the range between 200-800nm. Room temperature FTIR spectra were recorded as KBr pellet with an Alpha-Bruker FTIR spectrophotometer in the range of 4000-400cm⁻¹. Nuclear magnetic resonance spectra with different core viz., ¹H NMR, ¹³C NMR and ³¹P NMR were recorded in either DMSO-d₆ or CDCl₃ on Bruker ADVANCE III 500mHz spectrometer. The fluorescence spectra of the synthesised compound in ethanol were recorded on fluorescence spectrophotometer, FP-8500, JASCO. Mass spectroscopy was recorded on ES-FIGIEAN ionization mass spectrometer.

Fig. 3419.79(N chromatography using silica gel. Hexane

2.2.1. Materials
Solvents used were purified and dried according to the standard procedure.

2.2.2. Characterisation Methods
The UV-Visible spectra were recorded on an Alpha-Bruker UV spectrophotometer equipped in the range between 200-800nm. Room temperature FTIR spectra were recorded as KBr pellet with an Alpha-Bruker FTIR spectrophotometer in the range of 4000-400cm⁻¹. Nuclear magnetic resonance spectra with different core viz., ¹H NMR, ¹³C NMR and ³¹P NMR were recorded in either DMSO-d₆ or CDCl₃ on Bruker ADVANCE III 500mHz spectrometer. The fluorescence spectra of the synthesised compound in ethanol were recorded on fluorescence spectrophotometer, FP-8500, JASCO. Mass spectroscopy was recorded on ES-FIGIEAN ionization mass spectrometer.

Fig. 3419.79(N chromatography using silica gel. Hexane

2.2.1. Materials
Solvents used were purified and dried according to the standard procedure.

2.2.2. Characterisation Methods
The UV-Visible spectra were recorded on an Alpha-Bruker UV spectrophotometer equipped in the range between 200-800nm. Room temperature FTIR spectra were recorded as KBr pellet with an Alpha-Bruker FTIR spectrophotometer in the range of 4000-400cm⁻¹. Nuclear magnetic resonance spectra with different core viz., ¹H NMR, ¹³C NMR and ³¹P NMR were recorded in either DMSO-d₆ or CDCl₃ on Bruker ADVANCE III 500mHz spectrometer. The fluorescence spectra of the synthesised compound in ethanol were recorded on fluorescence spectrophotometer, FP-8500, JASCO. Mass spectroscopy was recorded on ES-FIGIEAN ionization mass spectrometer.

Fig. 3419.79(N chromatography using silica gel. Hexane

2.2.1. Materials
Solvents used were purified and dried according to the standard procedure.

2.2.2. Characterisation Methods
The UV-Visible spectra were recorded on an Alpha-Bruker UV spectrophotometer equipped in the range between 200-800nm. Room temperature FTIR spectra were recorded as KBr pellet with an Alpha-Bruker FTIR spectrophotometer in the range of 4000-400cm⁻¹. Nuclear magnetic resonance spectra with different core viz., ¹H NMR, ¹³C NMR and ³¹P NMR were recorded in either DMSO-d₆ or CDCl₃ on Bruker ADVANCE III 500mHz spectrometer. The fluorescence spectra of the synthesised compound in ethanol were recorded on fluorescence spectrophotometer, FP-8500, JASCO. Mass spectroscopy was recorded on ES-FIGIEAN ionization mass spectrometer.
2.4. Synthesis of 3-bromomethylquinoxalin-2-one

3-bromomethylquinoxalin-2-one (1.6g, 0.01mol) and NBS (1.80g, 0.01mol) were refluxed overnight in 30ml ccl₄ containing 0.08g (0.0003 mol) benzoyl peroxide. The byproduct NBS was removed by filtration. The reaction medium was washed with ccl₄ and the solvent evaporated. (Red crystals, Yield: 71%) m.p. 118-120 °C (S. Fig. 4) FT-IR (KBr, cm⁻¹): 2924.09, 3053.32 (C-H, st), 3439.08 (N-H, st) 1674.21 (C=O, st) 1475.54 (C=N, st) 1379.10 (C-N, st) 763.81 (C-Br, st) (S. Fig. 5) ¹H-NMR (DMSO, ppm): 4.6 δ(2H, s) 7.8 δ (1H, d) 7.4 δ (2H, m) 7.2 δ (1H, m) 8.7 δ (1H, s); (Fig. 2)

2.5. 3-triphenylphosphonium 3-bromomethylquinoxalin-2-one

3-bromomethyl-quinoxalin-2-one (0.24g, 1mmol) and triphenylphosphate (0.26g, 1mmol) was dissolved together in acetonitrile(20ml). The solution was stirred overnight at 40°C. The resulting precipitate was recrystallized from toluene-methanol mixture (2:1) to yield brown sticky phosphonium ylide compound. (S. Fig. 6) FT-IR (KBr, cm⁻¹): 2924.09, 3057.17 (C-H, st), 3446.79 (N-H, st) 1710.86 (C=O, st), 1436.97 (C=N, st), 1311.59 (C-N, st), 858.32 (C-Br, st), 721.38 (C-P, st), 540.07 (P-Br, st) (S. Fig. 7) ³¹P-NMR (DMSO, ppm): 2.540δ (2H, s), 7.390δ (6H, m), 7.459δ (6H, m), 7.493δ (3H, m), 7.591 δ (2H, m), 7.619 δ (2H, m) (Fig. 2) ³¹P-NMR (DMSO, ppm): 25.6968 (1P, s)

2.6. Synthesis of vinyl benzaldehyde capped quinoxaline derivative.

The vinyl benzaldehyde capped quinoxaline derivative were prepared from the corresponding phosphonium salt using the well-known Wittig reaction[16-17]. The phosphonium salt (0.50g, 1mmol) and terephthalic acid-oxaldehyde (0.135g, 1mmol) were dissolved in a mixture of absolute ethanol and dry chloroform (12ml, 3+1 v/v) under N₂ atmosphere. The reaction medium was washed with ccl₄ in 30ml ccl₄ and the solvent evaporated. (Red crystals, Yield: 71%; after 24 hours)

3. Results and discussion

3.1. Synthesis and Characterisation

The quinoxaline derivatives have been prepared by Condensation of o-phenylene diamine with 1,2-dicarbonyl compound in acetic acid at 80°C resulted into 3-methylquinolin-2-one. The structure of the synthesised compound was confirmed by FTIR, ¹H NMR and MASS spectral techniques. Although water is a desirable solvent for chemical reactions for many reasons like, cost, safety and environmental impacts, use of water in our reactions have generated moderate yields only (~25% after 24 hours). However, the reaction holds goods with acidic medium. (Scheme 1)

The vinyl benzaldehyde capped quinoxaline derivatives have been prepared similar to our earlier report[17,24,25] The scheme of the entire reaction have been listed in scheme 2. The reaction was carried out in 4 stages.

Stage 1 – Condensation reaction between diamino and diketone compound
Stage 2 – Bromination of 3-methyl-quinolin-2-one with NBS in ccl₄
Stage 3 – Formation of phosphonium ylide compound
Stage 4 – VB-QUI, the target compound obtained by the reaction between phosphonium ylide with terephthaldehyde.

Formation of various stages of compounds were confirmed using spectral studies viz., UV, FTIR, ¹H, ¹³C and ³¹P NMR, MASS spectroscopy.

Fig. 1 summarised the UV spectral details of 3-methylquinolin-2-one. From the Figure 1 the n-π* and π-π* transitions were observed at 300-350nm and 200-250nm respectively, which implies the C≡N, C≡O and C=C in the 3-methylquinolin-2-one moiety. FTIR spectrum of 3-methylquinolin-2-one has shown in Supplementary Fig. 1. The peak at 3008.95, 2914.44, 2848.86cm⁻¹ were attributed to aromatic and aliphatic C-H stretching frequency. The peak at 3419.79 corresponds to N-H stretching frequency. The peak at 1570.06 cm⁻¹ implies the C≡N in the phenyl ring. The peak at 1664.57cm⁻¹ for C=O stretching frequency. The bending frequency at 1381.03cm⁻¹, 754.17cm⁻¹ reveals that C-N, C-H frequency respectively. The ¹H NMR spectrum of 3-methylquinolin-2-one have displayed in Supplementary Fig. 2 The doublet of doublet at 7.261-7.362 ppm, 7.493-7.591 ppm for aromatic protons and 2.631 ppm signal for aliphatic protons. Supplementary Fig. 3 shows...
the MASS spectrum of 3-methylquinoxalin-2-one. From the spectra molecular ion peak observed at 159.2 this value agreed well with the theoretical value.

The stage 2 bromo methylated quinoxaline derivative was confirmed by FTIR, $^1$H NMR spectroscopy. Supplementary Fig. 4 shows the FTIR spectrum of 3-bromomethyl-quinoxalin-2-one. It displays transmittance peak at 763.81 cm$^{-1}$ for functional group C-Br stretching and 2858.51- 3157.47 cm$^{-1}$ stretching frequency for phenyl nucleus. The peak at 1674.21 cm$^{-1}$ for C=O stretching frequency. Supplementary Fig. 5 depicts the $^1$H NMR spectrum of 3-bromomethyl-quinoxalin-2-one. According to this spectrum, the bromomethylated proton signal observed at downfield region at 4.6 ppm may be due to electronegative bromo group attached in the methyl proton and signal at 8.7 ppm for N-H proton and signal appeared at 7.2-7.8 ppm for aromatic protons.

The stage 3 Phosphonium ylide was obtained from bromomethylated quinoxaline. Supplementary Fig. 6 summarised the FTIR spectrum of phosphonium salt. FTIR information for the formation of the phosphonium salt were confirmed by the appearance of P-Br, C-P stretching frequency at 534.28 cm$^{-1}$, 707.88 cm$^{-1}$. Supplementary Fig. 7 depicts the $^1$H NMR spectrum of phosphonium salt it displays the aromatic signals around 7.260-7.639 ppm and methylene proton signal at 2.515-2.602 ppm. Fig. 2 shows the $^{31}$P NMR spectrum of phosphonium compound. They displayed singlet signal at 25.698 ppm due to single phosphorous appeared in the compound.

The stage 4 vinyl benzaldehyde containing quinoxaline derivative has been obtained from phosphonium salt. Supplementary Fig. 8 summarised FTIR spectrum of QUI-PPV. It displays week transmittance peak at 2918 cm$^{-1}$ for C-H stretching frequency. The peak at 3392cm$^{-1}$ due to N-H stretching and peak at 1588cm$^{-1}$, 1689 cm$^{-1}$, 1429cm$^{-1}$ for C=N, C=O, C=C stretching frequency. The C-N stretching frequency appeared at 1347cm$^{-1}$. The $^1$H NMR spectrum of VB-QUI compound have shown in Supplementary Fig. 9. They displayed multiplets at 7.167-8.199 ppm attributable to aromatic protons, the signal at 10.02pmpp for N-H proton and 3.946, 5.559 ppm due to olefinic protons and 1.22 ppm attributable to aliphatic protons. Supplementary Fig. 10 show the $^{13}$C NMR spectrum of VB-QUI compound. They displayed signal at 191.00 ppm for carbonyl carbon and signal at 126.74 – 127.97 ppm due to vinyl carbons, further the signals appeared at 129.00-150.98 ppm for aromatic ring carbons. Supplementary Fig. 11 summarised the UV spectrum of VB-QUI compound. From the Figure $\lambda_{\text{max}}$ was observed at 200-252 nm which implies $\pi-\pi^*$ transition. The molecular weight was found from the GC-MASS spectrum shown in Fig 3. The molecular ion peak was observed at 277.24 found to be agreed well with the theoretical value.

3.2. Photoluminescent properties

Supplementary Fig. 10 displays the uv-vis absorption of the VB-QUI compound in the ethanol solution. The uv-vis absorption spectra of the solution exhibited the band around 200-250nm may be due to the $\pi-\pi^*$ electronic transition associated with the $\pi$-conjugation in the compound. Fig. 4 displays PL spectra of VB-QUI compound in the ethanol solution. In the PL spectra the compound showed a strong green emission approximately 470nm.

3.3. Anti-bacterial activity

The anti-bacterial activity of the synthesised vinyl benzaldehyde substituted quinoxaline was evaluated using two-Gram positive (Bacillus subtilis and Staphylococcus aureus) and two-Gram negative (Escherichia coli and Pseudomonas auroginosa) bacteria. Ampicillin is used as positive control. The MIC values of the compound were determined by broth dilution method[26]. Among the tested micro-organism the VB-QUI compound exhibited the best antibacterial activity, with a MIC value of 0.12mg except Gram-negative Pseudomonas auroginosa bacteria.

III. CONCLUSIONS

The vinyl benzaldehyde capped quinoxaline derivative compound was synthesised through Wittig reaction using Phosphonium salt and terephthaldehyde. The resulting compound was characterised by UV, FTIR. $^1$H, $^{13}$C, $^{31}$P NMR and GC-MASS spectral studies. The vinyl benzaldehyde introduced into the quinoxaline derivatives in the conjugation unit showed strong green emission at 470nm in the Photoluminescence spectra. Anti-bacterial activities of the synthesised compound were studied using Gram positive and Gram negative bacteria. In comparison with positive control ampicillin, the compound show relatively good anti-bacterial activity against tested micro-organism except Gram-negative Pseudomonas auroginosa bacteria.

ACKNOWLEDGEMENT

We gratefully acknowledge Muthurangam Govt Arts College (Autonomous) for providing laboratory facilities. We thank SIF-VIT and SAIF-IIT madras for recording spectral data.
REFERENCES


Table 1: Antibacterial study of the vinyl benzaldehyde capped quinoxaline derivative compound

<table>
<thead>
<tr>
<th>Compound</th>
<th>Bacillus subtilis</th>
<th>Staphylococcus aureus</th>
<th>Escherichia Coli</th>
<th>Pseudomonas aeruginosa</th>
</tr>
</thead>
<tbody>
<tr>
<td>PVB-QUI</td>
<td>0.12mg</td>
<td>0.12mg</td>
<td>0.125mg</td>
<td>0.25mg</td>
</tr>
</tbody>
</table>

Table 2: MIC (Minimum inhibitory concentration) values of VB-QUI compound
A Focus towards p-vinyl benzaldehyde containing quinoxaline – Photoluminescence...

Stage I: 3-methylquinoxalin-2-one

Figure 1: UV spectrum of 3-methylquinoxalin-2-one

Supplementary Figure 1: FTIR spectrum of 3-methylquinoxalin-2-one
A Focus towards p-vinyl benzaldehyde containing quinoxaline – Photoluminescence...

Supplementary Figure 2: $^1$H NMR spectrum of 3-methylquinoxalin-2-one

Supplementary Figure 3: MASS spectrum of 3-methylquinoxalin-2-one

Stage II: 3-bromomethyl-quinoxalin-2-one

Supplementary Figure 4: FTIR spectrum of 3-bromomethyl-quinoxalin-2-one
Supplementary Figure 5: $^1$H NMR spectrum of 3-bromomethyl-quinoxalin-2-one

Stage III: 3-triphenylphosphonium-bromomethylquinoxalin-2-one

Supplementary Figure 6: FTIR spectrum of 3-triphenylphosphonium-bromomethylquinoxalin-2-one

Supplementary Figure 7: $^1$H NMR spectrum of 3-triphenylphosphonium-bromomethylquinoxalin-2-one
A Focus towards p-vinyl benzaldehyde containing quinoxaline – Photoluminescence...

National Conference on Emerging Trends & Future Challenges in Chemical Sciences

Figure 2: $^{31}$P NMR spectrum of 3-triphenylphosphonium-bromomethylquinoxalin-2-one

Stage IV: 1,2-dihydro-2-oxoquinoxalin-3-vinyl benzaldehyde

Supplementary Figure 8: FTIR spectrum of 1,2-dihydro-2-oxoquinoxalin-3-vinyl benzaldehyde

Supplementary Figure 9: $^1$H NMR spectrum of 1,2-dihydro-2-oxoquinoxalin-3-vinyl benzaldehyde.
A Focus towards p-vinyl benzaldehyde containing quinoxaline – Photoluminescence...

Supplementary Figure 10: $^{13}$C NMR spectrum of 1,2-dihydro-2-oxoquinoxalin-3-vinyl benzaldehyde.

Figure 3: GC MASS spectrum of 1,2-dihydro-2-oxoquinoxalin-3-vinyl benzaldehyde.

Supplementary Figure 11: UV spectrum of 1,2-dihydro-2-oxoquinoxalin-3-vinyl benzaldehyde.
A Focus towards p-vinyl benzaldehyde containing quinoxaline – Photoluminescence...

Figure 4: PL spectrum of 1,2-dihydro-2-oxoquinoxalin-3-vinyl benzaldehyde.