

Synthesis, characterization and evaluation of *in vitro* antioxidant and anti-inflammatory activity of 2-(4-oxo-2-phenylquinazolin-3(4H)-yl) substituted acetic acids

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Abstract

Quinazolinone nucleus is a very attractive and useful scaffold in medicinal chemistry. It had been a pharmacophore in a wide variety of biologically active compounds. Peptides are key regulators in cellular and intercellular physiological responses and possess enormous promise for the treatment of pathological conditions. Prompted by therapeutic importance of quinazolinones and peptide derivatives, these two vital moieties were combined together into a single molecule by varying the different amino acids were synthesized, characterized and evaluated for anti-inflammatory and antimicrobial activities.

Keywords: - Amino acids,antioxidant, anti-inflammatory, benzoxizanone, peptides,quinazolinone

1. Introduction

Heterocyclic chemistry comprises at least half of all organic chemistry research worldwide. In particular, heterocyclic structures form the basis of many pharmaceutical, agrochemical and veterinary products. Among a wide variety of nitrogen heterocycles that have been explored for developing pharmaceutically, quinazolinone plays an important role in medicinal chemistry and subsequently have emerged as a pharmacophore. Quinazolinones are classes of fused heterocycles that are of considerable interest because of the diverse range of their biological activities such as farnesyltransferase, gastric H⁺/K⁺-ATPase and MAP kinase p38 inhibitory properties [1], anticancer[2-5], antiulcer[6], anti-tubercular[7], anti-bacterial and antifungal[8-11], anti-HIV[12], CNS depressant[13], anticonvulsant[14], antihelminthic[15], analgesic and anti-inflammatory[16-18], antihypertensive[19], antidiabetic[20] and anti-oxidant activities[21].

Peptides are active regulators and information breakers and possess enormous promise for the treatment of pathological conditions that make them interesting for drug discovery. Opioid peptide activity within the central nervous system (CNS) is of particular interest for the treatment of pain owing to the elevated potency of peptides and the centrally mediated actions of pain processes. They are the molecules of paramount importance in the fields of health care and nutrition controlling the numerous body processes and represent as promising drugs of the future [22]. Despite their potential, peptides have been of limited use as clinically viable drugs chiefly due to their undesired intrinsic properties. Therefore the challenge of this decade is to produce small molecules which mimic peptides and proteins. Incorporation of peptides into the aromatic and heterocyclic congeners results in compounds with potent bioactivities. Thus, keeping in mind the pharmacological potential of quinazolinones and peptides as well as taking advantage of biodegradability and biocompatibility of peptides, peptide derivatives of quinazolinone were prepared to increase therapeutic efficacy.(introduction of research article)Quinazolinone peptides were reported for their anti-inflammatory,antioxidant,anthelminthic,antibacterial and antifungal activities[23,24]. In view of their therapeutic importance, quinazolinone peptide derivatives were synthesized,characterized and evaluated for *in vitro* antioxidant and anti-inflammatory activities.

2. Materials and methods

Melting points were determined in an open capillary tube in Sigma Melting point apparatus and are uncorrected. Infrared Spectra of compounds were measured on a PE FTIR, in KBr disc and absorption bands expressed in cm⁻¹. ¹HNMR Spectra were recorded on a Bruker Avance dpx-200(at 200 MHz)Spectrometer with CDCl₃ as a solvent with tetramethylsilane(TMS) as an internal reference. Mass spectra were scanned on Jeol GCmate Mass spectrophotometer. Thin layer chromatography was carried out on silica gel to monitor the reactions and to check the purity of the compounds. All reagents were of the highest purity available commercially.Arginine, Histidine,Leucine,Glycine,Alanine,Asparagine, Glutamine, acetone, sodium hydroxide,N-benzoyl glycine,anthranilic acid and ethanol are procured from E.MERCK Ltd,Mumbai.Methionine and proline obtained from SISCO research laboratories PVT Ltd, Mumbai. The Eddy's Hot plate instrument is from Sigma,Chennai.

2.1. Chemistry

2.1.1 Synthesis of 2-phenyl-4H-benzo[d][1,3]oxazin-4-one: To a stirred solution of anthranilic acid (0.05mol) in pyridine (60ml), benzoyl chloride (0.05mol) was added drop wise, maintaining the temperature 0-5°C for 1hr. The reaction mixture was stirred for another 2hrs at room temperature until the solid product was formed. The reaction mixture was neutralized with saturated sodium bicarbonate solution and pale yellow solid which separated was filtered, washed with water, and recrystallized from ethanol. Yield-85%, M.P- 113-115°C (Lit.113-115°C)

2.1.2 General Method of Synthesis of 2-(4-oxo-2-phenylquinazolin-3(4H)-yl)substituted acetic acids (B1-B15): Glycine (0.01mol) in glacial acetic acid (10ml) and dry pyridine(10ml) was added to 2-phenyl-benzoxazine-4-one (0.01mol) and refluxed for 4hrs. The obtained reaction mixture was poured in to crushed ice and left overnight. The solid was filtered, washed with cold water, and re-crystallized from ethanol to obtain 2-(4-oxo-2-phenylquinazolin-3(4H)-yl) acetic acid (B1). Compounds B2-B15 were synthesized by the above mentioned procedure by condensing 2-phenyl-4H-benzo[d][1,3]oxazin-4-one with different amino acids. The scheme for synthesis is represented in Fig.1

2.2. Pharmacological activities

2.2.1. *In vitro* antioxidant activity

2.2.1.1. Interaction with stable free radical DPPH: Stable free radical species such as 1, 1-diphenyl-2-picrylhydrazyl (DPPH) is often used for the evaluation of the general radical scavenging capabilities of various antioxidants. DPPH, a paramagnetic compound with an odd electron, shows strong absorption band at 517nm. The absorbance decreases as a result of colour change from purple to yellow due to the scavenging of free radical by anti-oxidants through donation of hydrogen to form the stable DPPH-H molecule.

Solutions of various drugs at 100 µM concentration were added to 100 µM DPPH in 95% ethanol and tubes were kept at an ambient temperature for 20 minutes and absorbance was measured at 517nm. The drug concentration having 50% radical inhibition activity (IC₅₀) was calculated from the graph of % of free radical scavenging activity against the drug concentration. (Note: DPPH solution should be made freshly and should be kept in dark.)[25,26].

$$\text{Inhibition(\%)} = \frac{(\text{Control} - \text{Test})}{\text{Control}} \times 100$$

2.2.1.2 Scavenging of nitric oxide radical : Nitric oxide was generated from sodium nitroprusside and measured by Griess' reaction. Sodium nitroprusside (5mM) in standard phosphate buffer pH 7.4 were incubated with 100 µM concentrations of drug dissolved in a suitable solvent (dioxan/methanol) and the tubes were incubated at 25°C for 5hrs. Control experiment was conducted in an identical manner without test compound but with equal amount of solvent. After 5hrs, 0.5ml of incubation solution was removed and diluted with 0.5ml of Griess' reagent. The absorbance of the chromophore formed during diazotization of nitrite with sulphanilamide and its subsequent coupling with N-naphthyl ethylene diamine was read at 546nm. Composition of Griess reagent is Sulphanilamide(1%) +N-Naphthylethylene diamine(0.1%)+Orthophosphoric acid(2%) in 100ml Distilled water [27,28].

$$\text{Inhibition(\%)} = \frac{(\text{Control} - \text{Test})}{\text{Control}} \times 100$$

2.2.2. Acute inflammatory model-carrageenan induced paw edema assay: Male and female albino rats (150-200g) were used. The animals were fed with commercial feed pellets and were given water ad libitum. Carrageenan was obtained from s d fine chem. Ltd. Paw edema was measured by UGO BASILE 7140 Plethysmometer.

The rats were divided into 15 groups, one group consisting of six animals served as control, while the other groups of five animals each received the test compounds and standard drug. The rats were administered orally with test compounds (100 mg/kg), 100 mg/kg Diclofenac sodium or Ibuprofen (positive controls) or 10 ml/kg 0.5% sodium carboxy methyl cellulose (vehicle controls) one hour before injection of 0.05ml of 1% suspension of Carrageenan into the sub plantar region of the rat hind paw.

The volume of the injected paw was measured by water displacement in a digital plethysmograph immediately after carrageenan injection. The paw volume was again measured after 3 hours. A mark was made at the lateral maleolous of the right paw and the foot was dipped to the same distance of the mark into the arm of plethysmograph. Average edema volumes

for test compound treated and positive control rats were compared statistically with those of the vehicle control animals and expressed as percent edema inhibition, which is calculated using the formula. (Otterness I et al., 1985).

$$\text{Percentage edema inhibition} = 100 (1 - V_t/V_c)$$

Where, V_t = volume of edema in treated group, V_c = volume of the edema in the control group

Statistical analysis: Statistical analysis of the differences observed between control and treated groups were carried out using ANOVA. P value <0.05 was considered significant. Dunnett's post ANOVA has been done.

3. Results and Discussion

3.1. Chemistry

Fifteen compounds (B1-B15) were synthesized with the yields generally ranging from 70-90%. (B8) and (B9) derivatives were obtained at lowest yield 45-50%. The physical data such as melting points and yields are given in the Table I. The quinazolinone derivatives of the present study were characterized by UV, IR, ^1H NMR, and mass spectral analysis. The UV absorption peaks were observed in the region of the 230-320nm.

The IR spectra of all compounds displayed characteristic bands IR (KBr): ν 3365 (O-Hstr, Ar-OH), 3295-2505 (O-Hstr, COOH), 3072-3066, 3052 (Ar-Hstr), 2967, 2875 (C-Hstr, CH₃), 1702 (C=Ostr, COOH), 1669 (C=Ostr, ring), 1589, 1575, 1425, 1417 (skeletal bands), 1405 (O-Hdef, COOH), 875, 836, 760, 752, 696 (C-Hdef, Ar), strong bands at cm^{-1} due to C=O (in ring) stretching, cm^{-1} 3000-3085 due to C-H (Ar-H) stretching, cm^{-1} 1400-1587 due to C=C stretching (Ar).

^1H NMR spectra were taken for all the compounds which also supported the structures assigned. All the compounds displayed multiplets in the region of δ 6.6-8.3 due to aromatic hydrogen (Ar-H) in additional Compound B2 displayed a doublet in the region of δ 1.15-1.22 due to $-\text{CH}_3$ protons and quartet in the region of δ 4.3-4.5 due to $-\text{CH}$ -proton and a singlet in the region of δ 11.3-11.5 due to COO₂. Compound B4 displayed a doublet in the region of δ 1.2 -1.3 due to SH, a triplet at 2.5-2.8 due to $-\text{CH}_2$ protons another triplet in the region of 4-4.3 due to $-\text{CH}$ proton Compound B8 displayed a doublet in the region of δ 2.7-2.9 due to O-H proton (CH-OH). The structure of the compounds was also assigned by mass spectral analysis which showed (M⁺) peaks of the compounds.

3.2. In vitro antioxidant activity

3.2.1. Reduction of DPPH: All the compounds (B1-B15) were screened for reduction of DPPH. The highest activity of 84.9% was exhibited by B15, with guanidine group moiety. The compound with simplest amino acid, glycine (B1) showed only 35.6% of activity. When the alkyl chain was increased, there was no change in activity. B2, B3, B4 exhibited 37.8, 39.6 and 42.2% of activities respectively. When polar side chain amino acids were introduced, such as hydroxyl containing serine, threonine and tyrosine and sulfhydryl containing cysteine there was an increase in activity observed. B5, B7, B8, B11 exhibited 79, 78.1, 76.2 & 82.4% activities respectively. The results are given in Table-II

3.2.2. Nitric oxide radical scavenging: All the compounds (B1-B15) were tested for the scavenging of the nitric oxide free radical. Interestingly the compounds exhibited the same pattern of activity as in case of DPPH reduction. The compound with simplest amino acid, glycine (B1) showed only 31.2% of activity. When the alkyl chain was increased, there was no change in activity. B2, B3, B4 exhibited 33.1, 32.9 and 38.5% of activities respectively. When polar side chain amino acids were introduced, such as hydroxyl containing serine, threonine and tyrosine and sulfhydryl containing cysteine there was an increase in activity observed. B5, B7, B8, B11 exhibited 68.2, 61.6, 72.3 & 79.1% activities respectively. The highest activity of 79.7% was exhibited by B15, with guanidine group moiety. The results are given in Table-II

3.3. Anti-inflammatory activity

Among the tested compounds, the compound B1 exhibited 45.2% edema inhibition, when the compounds were prepared using non polar aliphatic and aromatic amino acids there was no much difference in the activity (43-50%). The compounds with polar aliphatic and polar aromatic amino acids there was a considerable rise in potency (57-69.8%). The compound B16 with arginine exhibited the highest anti-inflammatory activity of 71.6%.

4. Conclusion

The synthetic procedure was easier and %yields of the compounds were fairly good(78-92%). All the compounds exhibited moderate to potent and significant antioxidant and anti-inflammatory activities. Compound B15 showed extreme significant activity. The presence of essential structural features of good antioxidants in the synthesized compounds satisfy the criteria and thus proved to be potent. The ongoing development of these strategies provides promise that quinazolinone peptide drugs may be useful for the treatment of variety of diseases related to free radicals and inflammation.

Acknowledgement: We are thankful to IPT,SPMVV,Tirupati for providing the necessary facilities ,IIT madras for spectral data and UGC-New Delhi for granting the RFSMS.

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Fig.1 : General method for preparation of 2-(4-oxo-2-phenylquinazolin-3(4H)-yl)substituted acetic acids (B1-B15)

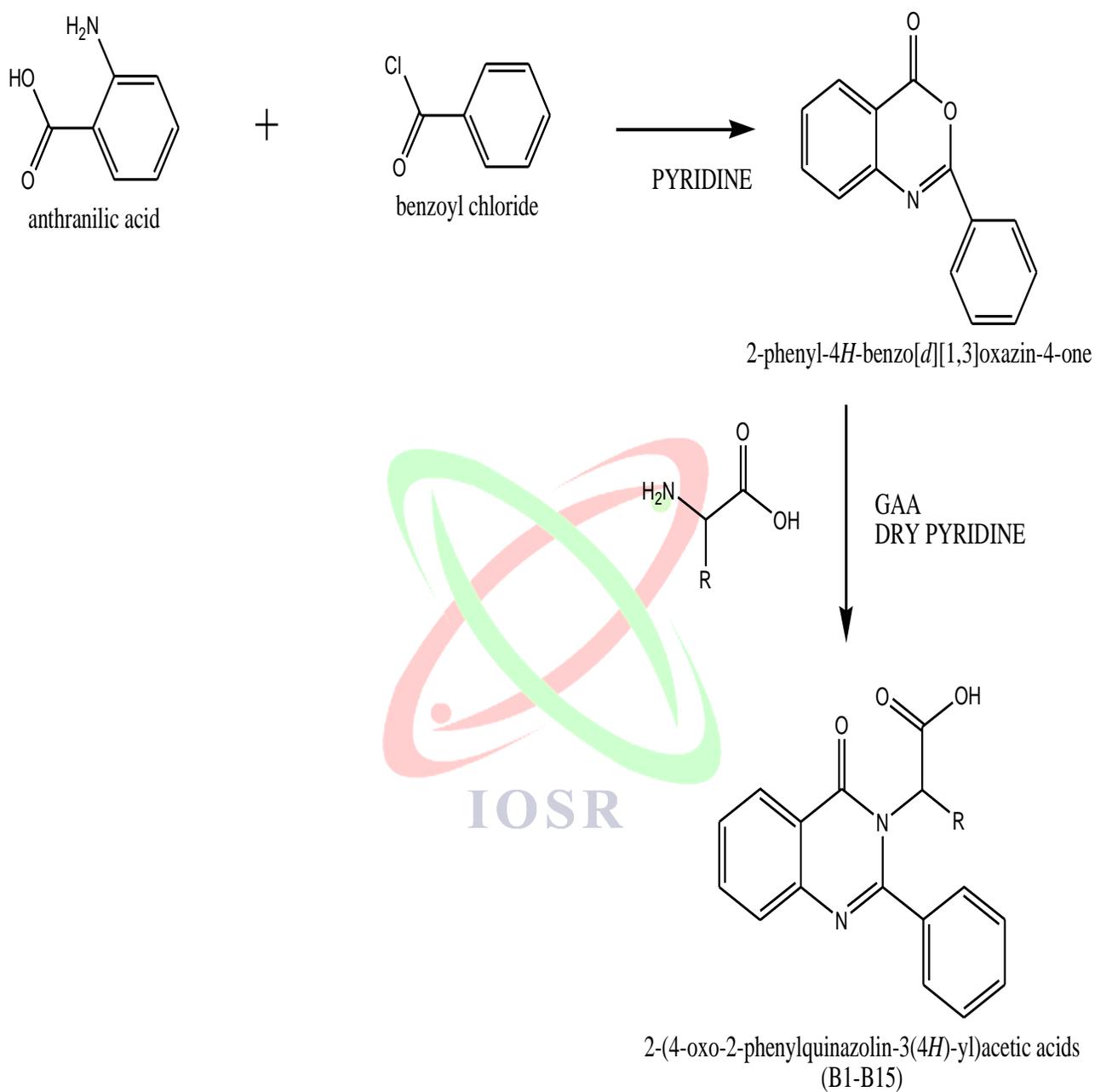
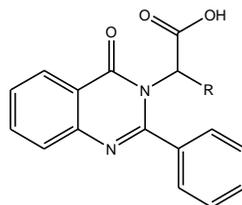


Table I: Physical data of 2-(4-oxo-2-phenylquinazolin-3(4H)-yl)substituted acetic acids (B1-B15)



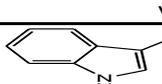
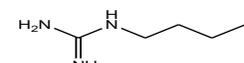
Compd	R	M.P., °C	Yield %
1	-H	134-136	58
2	-CH ₃	143-146	79
3	-CH ₂ -CH-(CH ₃) ₂	120-122	70
4	- CH(CH ₃)-CH ₂ -CH ₃	190-193	93
5	-CH ₂ -SH	128-130	78
6	-CH ₂ -S-CH ₃	240-244	81
7	-CH ₂ OH	173-175	65
8	- CH(CH ₃)-OH	180-183	53
9	-CH ₂ -CONH ₂	130-132	55
10	-Ph	240-242	57
11	-Ph(4-OH)	240-242	73
12		243-245	71
13	-(CH ₂) ₂ -COOH	230-232	65
14		120-124	61
15		130-132	81

Table II: *In vitro* Antioxidant and Anti inflammatory activity of (B1-B15) by carrageenan induced rat paw edema assay

Compound	Edema volume after 3rd hr (Mean±SEM)	% Edema inhibition after 3hours	% Reduction of DPPH(100µM)	%Nitric oxide scavenging (100µM)
Control	0.653±0.053 ^a	-	-	-
B1	0.296± 0.008 ^a	45.2	35.6	31.2
B2	0.306± 0.012 ^a	43.3	37.8	33.1
B3	0.270± 0.020 ^a	49.2	39.6	32.9
B4	0.166 ±0.008 ^a	51.6	42.2	38.5
B5	0.230± 0.026 ^a	62.2	79	68.2
B6	0.266± 0.008 ^a	57.7	62	61.6
B7	0.200± 0.005 ^a	68.2	78.1	72.3
B8	0.160± 0.026 ^a	69.8	76.2	74.7
B9	0.216± 0.029 ^a	60.3	46.8	42.8
B10	0.308± 0.013 ^a	43.5	53.5	49.3
B11	0.232± 0.026 ^a	62.1	82.4	79.1
B12	0.166 ±0.008 ^a	51.6	63	52.4
B13	0.246 ±0.017 ^a	46.4	72	63.5
B14	0.273 ±0.014 ^a	49	61.2	59.7
B15	0.150± 0.041 ^a	71.6	84.9	79.7
Standard	0.103± 0.023 ^a (Diclofenac sodium)	83.6	85 (tocopherol)	81 (curcumin)